Low Range Test Tube Format Phosphate Test Kit



(0.5 to 5.0 ppm Phosphate)

Enzyme and Reagents for 25 Sample Analysis (Including Standards)

1 ppm PO_4 -P = 3.06 ppm PO_4 (ppm $PO_4 = (94.97/31)$ ppm PO_4 -P)

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Phosphate-P	0.16 − 1.6 ppm PO ₄ -P	5.26 – 52.65 μM PO ₄ -P
Phosphate	0.5 - 5.0 ppm PO₄	5.26 − 52.65 µM PO ₄

Purine Nucleoside Phosphorylase (PNP) catalyzes the conversion of the artificial substrate MESG in the presence of inorganic phosphate to release a purine base which absorbs at 360 nm. The purine base that is produced is equimolar to the phosphate, therefore A-360 nm measures phosphate content.

This kit is supplied with enough reagents for 25 total samples, *including the standards*. Please keep this in mind when planning assays.

Supplied in Test Kit:

		MESG in dry powder form– 0.6+ mg
		1.5 mL tube in foil pouch
		Purine Nucleoside Phosphorylase (PNP) freeze-dried – 6.25 units
		1.5 mL tube in foil pouch
		HEPES/MgCl ₂ buffer in 50mL tube
		Phosphate Standard (100 ppm Phosphate) in liquid form – 1+ mL
		Standard tubes -7 tubes (5 mL) for preparing Phosphate Standards
		Dilution tube – (15 mL) for preparing stock standard
Supplie	d by	User:
		10 ml graduated cylinder
		Variable pipettes (10 to 100 µl and 100 to 1000µl)
		Spectrophotometer capable of reading at 360 nm, with
		a UV compatible cuvette (approx. volume 2 ml)
		(25)13 x 100 mm test tubes (Clean and Phosphate-free)
		Timer (0 to 40 minutes) – a clock or stop watch is adequate
		HPLC H ₂ O
		Ice and Ice Bucket



Reagent Preparation

Step 1 Prepare Assay Buffer – Warm the assay buffer to room temperature before use. Store at 4°C - stable for at least 6 months. Store dark.

Step 2 Prepare 2mM MESG – Please follow these instructions carefully

- Remove tube from foil pouch and tap tube to settle contents Before opening, make sure the reagent is not stuck to the cap.
- 2. Add 1 mL of HPLC water.
- 3. Re-cap and mix well by inversion. Make sure all the powder is in solution.
- 4. Do not vortex or over shake.
- 5. Keep on ice until use.

Step 3 Reconstitute PNP – Remove enzyme from foil pouch and tap tube to settle contents. Transfer 1.25 mL HEPES/ MgCl₂ buffer to tube, re-cap, and invert tube several times to mix *DO NOT VORTEX*.

Let set for at least 5 minutes and mix several more times. Concentration is now 5 units/mL.

Step 4 Prepare Reaction Mixture – Prepare fresh each for day of use

- 10.25 mL Assay Buffer (200mM HEPES, 20mM MgCl₂ pH 7.6)
- 1000 μL 2mM MESG
- 1250 μL reconstituted PNP (5 units/mL)
- Store reaction mixture in amber (or other light-tight) container. Make sure reaction mixture
 is room temperature before performing assays.
- Store any unused assay mix at -20°C or colder. It is best to use up all the reaction mixture
 the same day only using extra frozen reaction mixture for back-up.



REAGENT NOTES

Assay Buffer - 200 mM HEPES, pH 7.6 w/ 20 mM MgCl₂

MESG - 2 mM MESG in 200mM HEPES pH 7.6 w/ 20 mM MgCl₂

Purine Nucleoside Phosphorylase (PNP) – 6.25 units/tube

Phosphate Standard – 1 vial of 100 ppm Phosphate

Standard Preparation

Transfer 1 ml of 100 ppm Phosphate Standard into 15 mL tube containing 9 mL HPLC water to make a 10 ppm Phosphate Standard. Use the 7 screw cap tubes (provided in kit) to prepare Phosphate Standards as shown in table below. Cap and mix the tubes by inversion before use.

Vol 10 ppm PO ₄ Standard	Volume HPLC water	Resulting Standard (ppm PO ₄)	Resulting Standard (ppm PO ₄ -P)	Resulting Standard (µM)
-	5 mL	0	0	0
0.25 mL	4.75 mL	0.5	0.16	5.26
0.5 mL	4.5 mL	1.0	0.33	10.53
1.0 mL	4.0 mL	2.0	0.65	21.06
1.5 mL	3.5 mL	3.0	0.98	31.59
2.0 mL	3.0 mL	4.0	1.31	42.12
2.5 mL	2.5 mL	5.0	1.63	52.65



^{*}Thaw and/or make fresh reagents each day.

^{*}For best results, store all unused reagents at -20°C or colder.

^{*}Allow reaction mixture to reach room temperature (22°C) before beginning assays.

Phosphate Assay Procedure

- 1. Pipette 500 μL reaction mixture into each test tube or UV-compatible cuvette.
- 2. Pipette 500 μL sample, standard, or HPLC H₂O into each tube or cuvette.
- 3. Cap cuvettes and mix by inversion or vortex if using test tubes.
- 4. Incubate for 20 minutes at RT (22°C), mixing reaction tubes every 5 minutes.
- 5. Blank Spectrophotometer with the assay buffer.
- 6. Read samples at A-360 after 20 minute incubation time.
- 7. Create a standard curve. Plot samples against standard curve to get phosphate results.

Example Standard Curve:



