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### NECi Protocol for Nitrate Analysis in Plant Tissues: Laboratory Methods

Much of the nitrate in plant tissues is held within the cell walls. In order to determine total nitrate it is necessary to break the cells open. There are many standard techniques in use, including freezing and crushing the tissue, grinding in a blender, boiling, and combinations of these techniques. Here is a simple boiling method we've used with our kits. The standard ratio is one gram of sample per 10 milliliters of water (or extraction buffer).

Many plant tissues - especially root and leaf vegetables - can have nitrate at very high levels. Carrots and corn stalks may have 500 ppm or more. You will need to dilute these samples before analyzing them with our Nitrate Test Kits. Dilute extracts 1:10 and 1:10 again (using water or buffer), and analyze both dilutions.

For Quantitative work, dry samples should be used. You will find instructions for drying samples to constant weight on the reverse of this sheet.

1. Prepare a set of containers that can withstand boiling. Suitable containers include 50 ml centrifuge tubes or 25 ml Erlenmeyer flasks. Label one for each of your samples.
2. Cut each sample into small pieces. Place **one gram** of each sample into its labeled container. Add **10 ml** of deionized water (or extraction buffer) to each container.
3. **Optional step:** Add 50 mg activated charcoal to each flask. This step is required for quantitative data at levels below 1.0 ppm nitrate-N (below 70  $\mu$ M nitrate; Low Range kits), and is helpful for samples with high sugar content.
4. Set the labeled containers in a rack in your water bath. If a water bath is not available, use a pan of water on a hot plate. Be sure the keep flasks or other containers from tipping over. Bring flask contents to boil, and boil for 20 minutes. Cool to lukewarm before proceeding.
5. Decant the liquid contents of each container into a new vessel. Collect as much residual liquid as possible.
6. Bring volume back up to the initial 10 ml volume by adding deionized water. Mix.
7. **Optional step:** Clarify by filtration, preferably using a syringe filter. This is helpful for quantitative work, and necessary when analyzing samples below 1.0 ppm.
8. Samples are ready to be analyzed. Samples can also be stored in  $-20^{\circ}\text{C}$  freezer until analysis. You will need 50 $\mu$ l sample for Standard range kits (10 $\mu$ l for microplate); use 500  $\mu$ l sample for Low Range kits (50  $\mu$ l for microplate).

**Proceed to Nitrate Analysis. See *Screening Method* for simplified instructions for School, Home or Farm nitrate measurement.**