

Standard Range Test Tube Format Phosphate Test Kit

(5.0 to 20.0 ppm Phosphate)



Enzyme and Reagents for 25 Sample Analysis (Including Standards)

$5 \text{ ppm PO}_4\text{-P} = 15.32 \text{ ppm PO}_4$ ($\text{ppm PO}_4 = (94.97/31) \text{ ppm PO}_4\text{-P}$)

Phosphate-P	1.63 – 6.53 ppm PO₄-P	52.65 – 210.59 μM PO₄
Phosphate	5.0 - 20.0 ppm PO₄	52.65 – 210.59 μM PO₄

Purine Nucleoside Phosphorylase (PNP) catalyzes the conversion of the artificial substrate MESG in the presence of inorganic phosphate to release a purine base which absorbs at 360 nm. The purine base that is produced is equimolar to the phosphate, therefore A-360 nm measures phosphate content.

This kit is supplied with enough reagents for 25 total samples, *including the standards*. Please keep this in mind when planning assays.

Supplied in Test Kit:

- MESG** in dry powder form– 0.6+ mg
1.5 mL tube in foil pouch
- Purine Nucleoside Phosphorylase (PNP)** freeze-dried – 1.25 units
1.5 mL tube in foil pouch
- HEPES/MgCl₂** buffer in 50mL tube
- Phosphate Standard (100 ppm Phosphate)** in liquid form – 1+ mL
- Standard tubes** –5 tubes (1.5 mL) for preparing Phosphate Standards
- Dilution tube** – (5 mL) for preparing stock standard

Supplied by User:

- 10 ml graduated cylinder**
- Variable pipettes** (10 to 100 μl and 100 to 1000μl)
- Test tube vortex-type mixer** or similar to mix contents of tubes
- Spectrophotometer** capable of reading at 360 nm ± 20 nm, with a UV compatible cuvette (approx. volume 2 ml)
- (25) 13 x 100 mm test tubes** (Clean and Phosphate-free)
- Timer** (0 to 40 minutes) – a clock or stop watch is adequate
- HPLC H₂O**
- Ice and Ice Bucket**

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Reagent Preparation

Step 1 Prepare Assay Buffer – Warm the assay buffer to room temperature before use. Store at 4°C between uses - stable for at least 6 months. Store dark.

Step 2 Prepare 2mM MESG – Please follow these instructions carefully

1. Remove tube from foil pouch and tap tube to settle contents. Before opening, make sure the reagent is not stuck to the cap.
2. Add 1 mL of HPLC water.
3. Re-cap and mix well by inversion. Make sure all the powder is in solution.
4. Do not vortex or over-shake.
5. Keep on ice during day of use.

Step 3 Reconstitute PNP – Remove enzyme from foil pouch and tap tube to settle contents. Transfer 1 mL HEPES/MgCl₂ buffer to tube re-cap, and invert tube several times to mix *DO NOT VORTEX*. Let set for at least 5 minutes and mix several more times. Concentration is now 1.25 units/mL.

Step 4 Prepare Reaction Mixture – Prepare fresh each day

- 20.5 mL Assay Buffer (200mM HEPES, 20mM MgCl₂ – pH 7.6)
- 1000 µL 2mM MESG
- 1000 µL reconstituted PNP (1.25 units/mL)
- Store reaction mixture in amber (or other light-tight) container. Make sure reaction mixture is room temperature before performing assays.
- Store any unused assay mix at -20°C or colder. It is best to use up all the reaction mixture the same day- only using extra frozen reaction mixture for back-up.

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REAGENT NOTES

Assay Buffer – 200mM HEPES, pH 7.6 w/ 20 mM MgCl₂

MESG – 2mM MESG in 200mM HEPES pH 7.6 w/ 20 mM MgCl₂

Purine Nucleoside Phosphorylase (PNP) – 1.25 units/tube

Phosphate Standard – 1 vial of 100 ppm Phosphate

**Thaw and/or make fresh reagents each day.*

**For best results, store all unused reagents at -20°C or colder.*

**Allow reaction mixture to reach room temperature (22°C) before beginning assays.*

Standard Preparation

Transfer **1 ml of 100 ppm Phosphate Standard** into 5 mL tube containing **4 mL HPLC water** to make a **20 ppm Phosphate Standard**. Use the snap cap tubes (provided in kit) to prepare Phosphate Standards as shown in table below. Cap and mix the tubes well before use.

Vol 20 ppm PO ₄ Standard	Volume HPLC water	Resulting Standard (ppm PO ₄)	Resulting Standard (ppm PO ₄ -P)	Resulting Standard (µM)
-	1.0 mL	0	0	0
0.25 mL	0.75 mL	5	1.63	52.65
0.5 mL	0.5 mL	10	3.26	105.30
0.75 mL	0.25 mL	15	4.90	157.94
1.0 mL	0 mL	20	6.53	210.59

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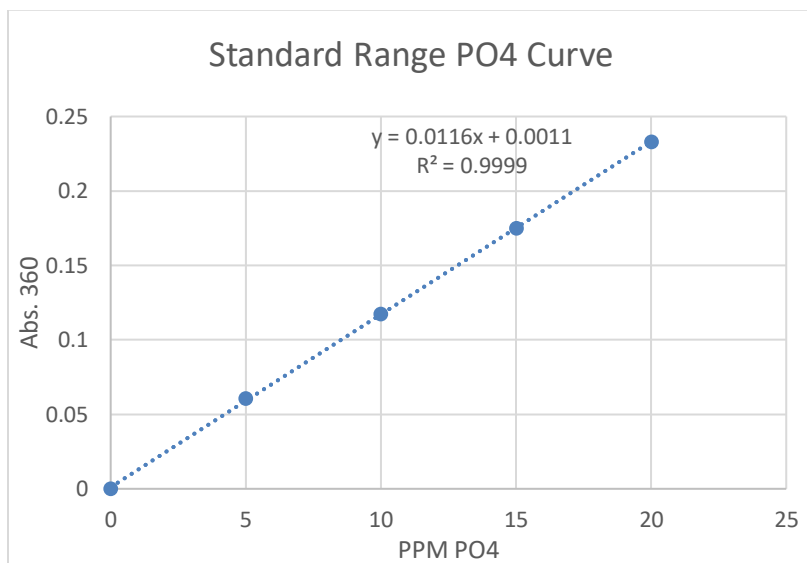
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Phosphate Assay Procedure

1. Pipette 900 μL reaction mixture into each test tube or UV-compatible cuvette.
2. Pipette 100 μL sample, standard, or HPLC H_2O into each tube or cuvette.
3. Cap cuvettes and mix by inversion or vortex if using test tubes.
4. Incubate for 20 minutes at RT (22°C), mixing reaction tubes every 5 minutes.
5. Blank Spectrophotometer with the assay buffer.
6. Read samples at A-360 after 20 minute incubation time.
7. Create a standard curve. Plot samples against standard curve to get phosphate results.

Example Standard Curve:



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