

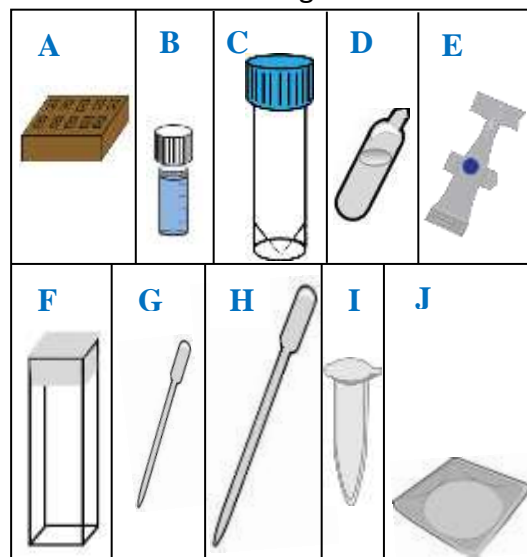
# Dry Forage Nitrate Test Kit Instructions

## Introduction

This kit contains everything needed to test dry forage samples for nitrate content. Provided in this kit is a **1000 ppm Nitrate-N standard (D)**, to use as a reference tool for analyzing samples and to ensure that the kit is working properly. Download the free “Nitrate Color Slider” application, available on Google Play and iTunes App, for easy analysis of your samples on mobile devices. A color chart is also provided on the back of this instruction sheet for sample analysis. These kits are based on validated laboratory methods, and will provide nitrate results after 15-20 minutes. Although results are not as precise as a testing lab, *you will get accurate nitrate content results* for making immediate decisions about feeding livestock or further testing.

### Kit Contents (per 5 samples):

- A.** 1 cardboard cuvette holder
- B.** 5 clear collection tubes with white screw caps holding 4.5 mL water
- C.** 1 50 mL tube with blue screw cap
- D.** 5 small squeeze bulbs (contains buffer)
- E.** 1 large squeeze bulb with blue dot (contains *nitrate standard*)
- F.** 6 square reaction cuvettes in sealed foil pouch (contains enzyme)
- G.** 5 small plastic pipettes
- H.** 5 large plastic pipettes
- I.** 6 snap-cap tubes of color reagent powder (in amber bag)
- J.** 1 large plastic pan (for placing ground up sample into)



### Materials you will need:

- Marker for labeling tubes and cuvette caps, pen for recording data
- Scissors (to remove tip from *snip off* squeeze bulb)
- Teaspoon to measure sample
- Kitchen blender to grind sample
- Distilled water to add to 50 mL tube

#### DO:

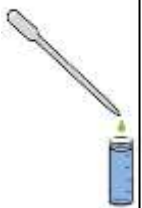
- ✓ Store this kit in a cool and dark place (below 72°F/22°C), refrigerate if possible
- ✓ Follow the directions of a Certified Crop Advisor for advice on plant sampling.
- ✓ Run tests and standard all at once to ensure accuracy
- ✓ Label collected sample tubes and reaction cuvettes accordingly
- ✓ Gently shake cuvettes several times throughout reaction without inverting them.

#### DO NOT:

- \*Open sealed packets or mix cuvette contents until ready to use
- \*Add more than *one drop* of sample water to reaction cuvette
- \*Compare samples to color chart before waiting at least 10 minutes
- \*Invert reaction cuvettes when mixing or get liquid in the cap portion before step 4

## Procedure

1.3

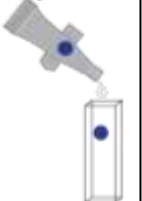


### Step 1 Sample Preparation

- 1) Grind a handful of sample until pieces are uniformly ¼" or less. Transfer to plastic pan.
- 2) Add distilled water to the black mark on the large tube (blue cap). Add 1 rounded tsp of sample to tube. Cap.
- 3) Shake tube for 1 minute and let settle for 5 minutes. Use one of the large pipettes to transfer 0.5mL of this extract to one of the collection tubes with white cap\*(1.3). Dump out and rinse large tube with distilled water.
- 4) Repeat above steps for other samples, using a clean pipette for each sample.

\*Be sure to label each sample tube cap and record the information regarding the samples on the data sheet (1-5, A-E, etc.)

2.3



### Step 2 Preparing the Reaction Cuvettes

- 1) Remove the 6 square reaction cuvettes from the foil pouch and place them in the cardboard cuvette holder, keeping the one with the blue dot separate from the others (this is for your **nitrate standard**).
- 2) Tap cuvettes to settle contents making sure to keep them upright to not lose contents.
- 3) Twist off the end of the squeeze bulb with the blue dot and empty the entire contents into the cuvette with the blue dot. This is the **nitrate standard**. (2.3)
- 4) Label the other cuvette caps with numbers, letters, or location to match sample tubes. Into each of the other 5 cuvettes, empty entire contents of one buffer squeeze bulb. (2.5)
- 5) Cap all cuvettes and mix gently. Do not invert (keep liquid out of cap). Step 3 must follow within 5 minutes.

2.5



### Step 3 Adding samples to Reaction Cuvettes

- 1) Within 5 minutes, pick up liquid from near the surface of your first sample with a small plastic pipette (3.1).
- 2) Transfer only one drop (approximately 50 µl) of this liquid to the corresponding cuvette (Sample 1 into cuvette 1, etc.) making sure not to touch the sides or surface of the liquid with the pipette. (3.2)
- 3) Repeat for the remaining 4 samples, making sure to use a new pipette for each unique sample.
- 4) Recap the cuvettes and mix by gently shaking side to side.
- 5) Let set for **at least 10 minutes**, mixing gently every few minutes.

3.1



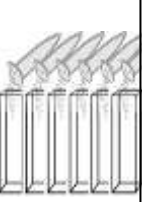
3.2



### Step 4 Color development

- 1) Tap each snap-cap tube to a hard surface to settle color reagent powder; open tubes.
- 2) Add color reagent powder to all reaction cuvettes, including the nitrate standard. (4.1)
- 3) Firmly seal the cuvettes with square caps and vigorously mix each cuvette rapidly.
- 4) Let cuvettes develop color for about 5 minutes, mixing them several times to dissolve most of the powder. Some powder may settle to the bottom of tube; this is okay.

4.1



### Step 5 Evaluating your results

After color development, compare your results within one hour using the **nitrate standard** and this chart. Use the QR code or visit our website to use our custom color slider app. The **nitrate standard** is set at 1000 ppm Nitrate-N (4500 ppm Nitrate). This is the point at which you should use caution for feeding to livestock.



Nitrate-N (ppm)

<350

350-1000

1000-2500

> 2500

Nitrate (ppm)

<1500

1500-5000

5000-10,000

>10,000

**Recommendation For Feeding**

Generally considered safe for all livestock

Limit to 50% for calves, pregnant, or lactating animals.

Limit to 25%-50% feed. Do not feed to pregnant animals.

Use Caution!  
Potentially Deadly!  
Do not feed as is.

Download our free Nitrate Color Slider app available on Google Play and iTunes for easy color matching of your samples for data. Visit [www.nitrate.com/mobile-apps](http://www.nitrate.com/mobile-apps) on your mobile device's web browser, or search "Nitrate Color Slider" in the app store. Check out NECi's Handheld Photometer and app for digital data on your mobile device!